

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**Applicants:** Leonard Harrison, et al**Examiner:** Unassigned**Serial No.:** Unassigned**Docket:** 10308B**Filed:** Herewith**Dated:** January 7, 2002**For:** IMMUNOREACTIVE AND
IMMUNOTHERAPEUTIC MOLECULESAssistant Commissioner for Patents
Washington, DC 20231**PRELIMINARY AMENDMENT**

Sir:

Kindly enter the following amendments prior to examining the above-captioned application.

IN THE SPECIFICATION:**Page 1, line 4,** please insert the following text:**--CROSS REFERENCE TO RELATED APPLICATION**

CERTIFICATE OF MAILING BY "EXPRESS MAIL"**"Express Mail" Mailing Label Number: EV 052766565 US****Date of Deposit: January 7, 2002**

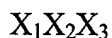
I hereby certify that this correspondence is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 C.F.R. §1.10 on the date indicated above and is addressed to the Assistant Commissioner of Patents and Trademarks, Washington, D.C. 20231.

Dated: January 7, 2002
Michelle Mustafa

The present application is a divisional application of U.S. Serial No. 08/663,272
filed on November 25, 1996.--

Please replace the paragraph beginning at Page 3, line 18, with the following rewritten paragraph:

--According to this preferred embodiment, there is provided a recombinant or
synthetic peptide or chemical equivalent thereof comprising the sequence:

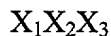


wherein

X₁ and X₃ may be the same or different and each is an amino acid sequence
comprising from 0 to 15 naturally or non-naturally occurring amino acid residues; X₂ is selected
from FFYTPKTRREAED and FWYIPPSLRTLED or a derivative or chemical equivalent thereof
and wherein said peptide is capable of reacting with T cells and modifying T-cell function when
incubated with cells from subjects with pre-clinical or clinical IDDM and determining reactivity by
an appropriate assay. Preferred cells include but are not limited PBMCs, anti-coagulated whole
blood or tissue biopsy cells and determining reactivity by an appropriate assay.--

Please replace the paragraph beginning at Page 10, line 6, with the following rewritten paragraph:

--Preferably the present invention contemplates a method of assaying the reactivity
of a subject to IDDM autoantigen said method comprising contacting a peptide or chemical
equivalent thereof comprising the formula



wherein:

X₁ and X₃ may be the same or different and each is an amino acid sequence comprising from 0 to
15 naturally or non-naturally occurring amino acid residues; X₂ is selected from
FFYTPKTRREAED and FWYIPPSLRTLED or a derivative or chemical equivalent thereof and
wherein said peptide is capable of reacting with T cells and modifying T-cell function when

incubated with cells from subjects with pre-clinical or clinical IDDM and determining reactivity by an appropriate assay. Preferably cells include but are not limited to peripheral blood mononuclear cells (PBMCs), anticoagulated whole blood and tissue biopsy cells.

Please replace the paragraph beginning at Page 11, line 6, with the following rewritten paragraph:

--BRIEF DESCRIPTION OF THE DRAWINGS--.

Please replace the paragraph beginning at Page 11, line 8, with the following rewritten paragraph:

--Figure 1 shows a comparison of the regions of similarity among mouse and human proinsulins and GADs (SEQ ID NOS:1-7). Similarities are boxed; identities within boxes are shaded. The C-terminus of the mature insulin B-chain and the pro-insulin cleavage site are indicated by the vertical line and arrow respectively.--

Please replace the paragraph beginning at Page 11, line 13, with the following rewritten paragraph:

--Figures 2A and 2B are graphical representations showing the level of cellular proliferation expressed as the delta score following the stimulation of peripheral blood mononuclear cells taken from IDDM at-risk (as described in Example 1) or control subjects with the following peptides in Figure 2A: human GAD65 (residues 506-518); human proinsulin (residues 24-36); irrelevant control peptide; or in Figure 2B tetanus toxoid (CSL Ltd., Melbourne, Australia).--

Please replace the paragraph beginning at Page 14, line 20, with the following rewritten paragraph:

--Blood was drawn from paired IDDM at-risk and HLA-DR matched controls at the same time (within 30 minutes) and processed similarly to reduce the effects of diurnal variation and handling artifacts. Peripheral blood mononuclear cells were isolated from heparinised whole blood by Ficoll-Paque (Pharmacia Biotech) density centrifugation, washed and resuspended in RPMI 1640 medium (Biosciences Pty Ltd) containing 20mM Hepes (CSL Ltd), 10^{-5} M 2-mercaptoethanol (BDH), penicillin (100U/ml), streptomycin (100 µg/ml) and 10% v/v autologous

plasma. Aliquots of 200 μ l (2×10^5 cells) were transferred into wells of a 96-well, round-bottomed plate (Falcon) and incubated in replicates of six with the following peptides as final concentrations of 10, 2, and 0.4 μ g/ml: human GAD65 (506-518), human proinsulin (24-36) (synthesized using an Applied Biosystems Model 431A synthesizer (ABI, Foster City, CA), and an irrelevant control peptide (CRFDPQFALTNIIVRK) SEQ ID NO:8 (Macromolecular Resources, Fort Collins, CO). Tetanus toxoid (CSL Ltd, Melbourne, Australia) at final concentrations of 1.8, 0.18 and 0.018 LfU/ml was used as a positive control. Twelve "autologous only" wells containing cells but without antigen were included as the background control. Plates were incubated at 37°C in a 5% v/v CO₂ humidified incubator for 6 days; 0.25 μ Ci of [³H]thymidine (ICN) was added to each well for the last 6 hours. The cells were then harvested onto glass fibre filters and incorporated radioactivity measured by beta-particle counting (Packard Model 2000 Liquid Scintillation Counter). The level of cellular proliferation was expressed as the delta score (DS=mean counts per minute (cpm) incorporated in the presence of antigen, minus the mean cpm of the "autologous only" wells).--

Please replace the paragraph beginning at Page 15, line 25, with the following rewritten paragraph:

--Reactivity to the proinsulin sequence was confined almost entirely to IDDM at-risk subjects, whereas some controls also responded to the GAD peptide (Table 2, Figures 2A and 2B). Both groups responded similarly to tetanus, and no subject reacted to the unrelated control peptide.

Please replace the paragraph beginning at Page 23, line 1, with the following rewritten paragraph:

--WHAT IS CLAIMED IS:--

IN THE CLAIMS

Please cancel claims 1-7 and 30-36 without prejudice.

Please amend the Claims as follows:

10. (Amended) The method according to Claim 8 wherein an appropriate assay includes proliferation assay, cytotoxic assays, cellular reactivity or combinations thereof.

19. (Amended) A composition comprising a peptide or chemical equivalent thereof comprising the formula:



wherein:

X₁ and X₃ may be the same or different and each is an amino acid sequence comprising from 0 to 40 naturally or non-naturally occurring amino acid residues; X₂ is any amino acid sequence of from 10 to 100 residues derived from, homologous to or contiguous within amino acids 506 to 518 inclusive or derivatives thereof of human GAD 65 or amino acids 24 to 36 inclusive or derivatives thereof of human proinsulin; and wherein said peptide molecule is capable of reacting with T cells and modifying T-cell function when incubated with cells from subjects having pre-clinical or clinical Insulin-Dependent Diabetes Mellitus (IDDM) to assay reactivity of a subject to IDDM autoantigen by contacting said peptide or its chemical equivalent to cells from said subject and determining reactivity by an appropriate assay.

20. (Amended) The composition according to claim 19 wherein the cells are selected from the group comprising PBMCs, anti-coagulated whole blood or tissue biopsy cells.

21. (Amended) The composition according to claim 19 wherein an appropriate assay includes proliferation assay, cytotoxic assays, cellular reactivity or combinations thereof.

22. (Amended) The composition according to claim 19 wherein X₂ comprises from 10 to 50 amino acid residues.

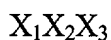
23. (Amended) The composition according to claim 22 wherein X₂ comprises from 10 to 30 amino acid residues.

24. (Amended) The composition according to claim 23 wherein X₂ comprises from 10 to 15 amino acid residues.

25. (Amended) The composition according to claim 24 wherein X₂ comprises the amino acid sequence: FFYTPKTRREAED.

26. (Amended) The composition according to claim 24 wherein X₂ comprises the amino acid sequence: FWYIPPSLRTLED.

27. (Amended) A composition comprising a peptide of chemical equivalent thereof comprising the formula:



wherein:

X₁ and X₃ may be the same or different and each is an amino acid sequence comprising from 0 to 15 naturally or non-naturally occurring amino acid residues; X₂ is selected from FFYTPKTRREAED and FWYIPPSLRTLED or a derivative or chemical equivalent thereof and wherein said peptide is capable of reacting with T cells and modifying T-cell function when incubated with cells from subjects with pre-clinical or clinical IDDM to assay reactivity of a subject to IDDM autoantigen by contacting said peptide or its chemical equivalent with cells from said subject and determining reactivity by a proliferation assay.

28. (Amended) The composition according to claim 27 wherein the cells are selected from the group comprising PBMCs, anti-coagulated whole blood or tissue biopsy cells.

29. (Amended) The composition according to claim 27 wherein an appropriate assay includes proliferation assay, cytotoxic assays, cellular reactivity or combinations thereof.

Please add the following new claims:

37. The method according to claim 8 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:1.

38. The method according to claim 8 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:2.

39. The composition according to claim 19 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:1.

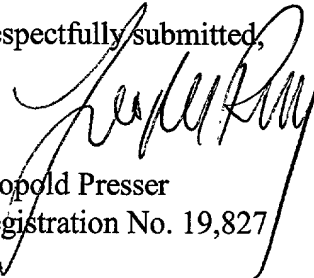
40. The composition according to Claim 19 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:2.

REMARKS

The present application is a divisional application of U.S. Serial No. 08/663,272 (the "272 application") filed on November 25, 1996. Applicants have amended the specification to include the sequence listing submitted on September 3, 1997 in the parent case (the '272 application). The specification has also been amended to identify sequences by sequence identification reference numbers. Other minor corrections have been made. No new matter is included in this submission.

In view of the foregoing amendment, it is respectfully submitted that the present application is in condition for allowance, which action is earnestly solicited.

Respectfully submitted,



Leopold Presser
Registration No. 19,827

SCULLY, SCOTT, MURPHY & PRESSER
400 Garden City Plaza
Garden City, New York 11530
(516) 742-4343

PIB/LP:dg

400 Garden City Plaza
Garden City, New York 11530
(516) 742-4343

Serial No.: Unassigned
Filed: Herewith
Docket: 10308B

VERSION WITH MARKINGS TO SHOW CHANGES MADE

IN THE SPECIFICATION:

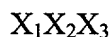
Page 1, line 4, please insert the following text:

--CROSS REFERENCE TO RELATED APPLICATION

The present application is a divisional application of U.S. Serial No. 08/663,272
filed on November 25, 1996.--

Please replace the paragraph beginning at Page 3, line 18, with the following rewritten paragraph:

--According to this preferred embodiment, there is provided a recombinant or
synthetic peptide or chemical equivalent thereof comprising the sequence:

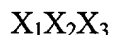


wherein

X₁ and [X₂] X₃ may be the same or different and each is an amino acid sequence
comprising from 0 to 15 naturally or non-naturally occurring amino acid residues; X₂ is selected
from FFYTPKTRREAED and FWYIPPSLRTLED or a derivative or chemical equivalent thereof
and wherein said peptide is capable of reacting with T cells and modifying T-cell function when
incubated with cells from subjects with pre-clinical or clinical IDDM and determining reactivity by
an appropriate assay. Preferred cells include but are not limited PBMCs, anti-coagulated whole
blood or tissue biopsy cells and determining reactivity by an appropriate assay.--

Please replace the paragraph beginning at Page 10, line 6, with the following rewritten paragraph:

--Preferably the present invention contemplates a method of assaying the reactivity
of a subject to IDDM autoantigen said method comprising contacting a peptide or chemical
equivalent thereof comprising the formula



wherein:

X_1 and $[X_2] X_3$ may be the same or different and each is an amino acid sequence comprising from 0 to 15 naturally or non-naturally occurring amino acid residues; X_2 is selected from FFYTPKTRREAED and FWYIPPSLRTLED or a derivative or chemical equivalent thereof and wherein said peptide is capable of reacting with T cells and modifying T-cell function when incubated with cells from subjects with pre-clinical or clinical IDDM and determining reactivity by an appropriate assay. Preferably cells include but are not limited to peripheral blood mononuclear cells (PBMCs), anticoagulated whole blood and tissue biopsy cells.

Please replace the paragraph beginning at Page 11, line 6, with the following rewritten paragraph:

[In the Figures:] --BRIEF DESCRIPTION OF THE DRAWINGS--.

Please replace the paragraph beginning at Page 11, line 8, with the following rewritten paragraph:

--Figure 1 shows a comparison of the regions of similarity among mouse and human proinsulins and GADs (SEQ ID NOS:1-7). Similarities are boxed; identities within boxes are shaded. The C-terminus of the mature insulin B-chain and the pro-insulin cleavage site are indicated by the vertical line and arrow respectively.--

Please replace the paragraph beginning at Page 11, line 13, with the following rewritten paragraph:

--[Figure 2 is a graphical representation] Figures 2A and 2B are graphical representations showing the level of cellular proliferation expressed as the delta score following the stimulation of peripheral blood mononuclear cells taken from IDDM at-risk (as described in Example 1) or control subjects with the following peptides in Figure 2A: human GAD65 (residues 506-518); human proinsulin (residues 24-36); irrelevant control peptide; or in Figure 2B tetanus toxoid (CSL Ltd., Melbourne, Australia).--

Please replace the paragraph beginning at Page 14, line 20, with the following rewritten paragraph:

--Blood was drawn from paired IDDM at-risk and HLA-DR matched controls at the same time (within 30 minutes) and processed similarly to reduce the effects of diurnal variation and handling artifacts. Peripheral blood mononuclear cells were isolated from heparinised whole blood by Ficoll-Paque (Pharmacia Biotech) density centrifugation, washed and resuspended in RPMI 1640 medium (Biosciences Pty Ltd) containing 20mM Hepes (CSL Ltd), 10^{-5} M 2-mercaptoethanol (BDH), penicillin (100U/ml), streptomycin (100 μ g/ml) and 10% v/v autologous plasma. Aliquots of 200 μ l (2×10^5 cells) were transferred into wells of a 96-well, round-bottomed plate (Falcon) and incubated in replicates of six with the following peptides as final concentrations of 10, 2, and 0.4 μ g/ml: human GAD65 (506-518), human proinsulin (24-36) (synthesized using an Applied Biosystems Model 431A synthesizer (ABI, Foster City, CA), and an irrelevant control peptide (CRFDPQFALTNIIVRK) SEQ ID NO:8 (Macromolecular Resources, Fort Collins, CO). Tetanus toxoid (CSL Ltd, Melbourne, Australia) at final concentrations of 1.8, 0.18 and 0.018 LfU/ml was used as a positive control. Twelve "autologous only" wells containing cells but without antigen were included as the background control. Plates were incubated at 37°C in a 5% v/v CO₂ humidified incubator for 6 days; 0.25 μ Ci of [³H]thymidine (ICN) was added to each well for the last 6hours. The cells were [than] then harvested onto glass fibre filters and incorporated radioactivity measured by beta-particle counting (Packard Model 2000 Liquid Scintillation Counter). The level of cellular proliferation was expressed as the delta score (DS=mean counts per minute (cpm) incorporated in the presence of antigen, minus the mean cpm of the "autologous only" wells).--

Please replace the paragraph beginning at Page 15, line 25, with the following rewritten paragraph:

--Reactivity to the proinsulin sequence was confined almost entirely to IDDM at-risk subjects, whereas some controls also responded to the GAD peptide (Table 2, Figures 2A and 2B) [(Table 2, Figure 2)]. Both groups responded similarly to tetanus, and no subject reacted to the unrelated control peptide.

Please replace the paragraph beginning at Page 23, line 1, with the following rewritten paragraph:

--[CLAIMS] WHAT IS CLAIMED IS:--

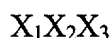
IN THE CLAIMS

Please cancel claims 1-7 and 30-36 without prejudice.

Please amend the Claims as follows:

10. (Amended) [A] The method according to Claim 8 [or 9] wherein an appropriate assay includes proliferation assay, cytotoxic assays [celular] cellular reactivity or combinations thereof.

19. (Amended) [Use of] A composition comprising a peptide or chemical equivalent thereof comprising the formula:



wherein:

X₁ and X₃ may be the same or different and each is an amino acid sequence comprising from 0 to 40 naturally or non-naturally occurring amino acid residues; X₂ is any amino acid sequence of from 10 to 100 residues derived from, homologous to or contiguous within amino acids 506 to 518 inclusive or derivatives thereof of human GAD 65 [and/or] or amino acids 24 to 36 inclusive or derivatives thereof of human proinsulin; and wherein said peptide molecule is capable of reacting with T cells and modifying T-cell function when incubated with cells from

subjects having pre-clinical or clinical Insulin-Dependent Diabetes Mellitus (IDDM) to assay reactivity of a subject to IDDM autoantigen by contacting said peptide or its chemical equivalent to cells from said subject and determining reactivity by an appropriate assay.

20. (Amended) [Use] The composition according to claim 19 wherein the cells are selected from the group comprising PBMCs, anti-coagulated whole blood [and/or] or tissue biopsy cells.

21. (Amended) [Use] The composition according to claim 19 [or 20] wherein an appropriate assay includes proliferation assay, cytotoxic assays, [celular] cellular reactivity or combinations thereof.

22. (Amended) [Use] The composition according to claim 19 wherein X₂ comprises from 10 to 50 amino acid residues.

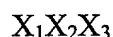
23. (Amended) [Use] The composition according to claim 22 wherein X₂ comprises from 10 to 30 amino acid residues.

24. (Amended) [Use] The composition according to claim 23 wherein X₂ comprises from 10 to 15 amino acid residues.

25. (Amended) [Use] The composition according to claim [19 or 20 or 21 or 22 or 23 or] 24 wherein X₂ comprises the amino acid sequence: FFYTPKTRREAED.

26. (Amended) [Use] The composition according to claim [19 or 20 or 21 or 22 or 23 or] 24 wherein X₂ comprises the amino acid sequence: FWYIPPSLRTLED.

27. (Amended) [Use of] A composition comprising a peptide of chemical equivalent thereof comprising the formula:



wherein:

X₁ and [X₂] X₃ may be the same or different and each is an amino acid sequence comprising from 0 to 15 naturally or non-naturally occurring amino acid residues; X₂ is selected from FFYTPKTRREAED and FWYIPPSLRTLED or a derivative or chemical equivalent thereof and wherein said peptide is capable of reacting with T cells and modifying T-cell function when incubated with cells from subjects with pre-clinical or clinical IDDM to assay reactivity of a subject to IDDM autoantigen by contacting said peptide or its chemical equivalent with cells from said subject and determining reactivity by a proliferation assay.

28. (Amended) [Use of a peptide or chemical equivalent] The composition according to claim 27 wherein the cells are selected from the group comprising PBMCs, anti-coagulated whole blood [and/or] or tissue biopsy cells.

29. (Amended) [Use of a peptide of chemical equivalent] The composition according to claim 27 [or 28] wherein an appropriate assay includes proliferation assay, cytotoxic assays, cellular reactivity or combinations thereof.

Please add the following new claims:

37. The method according to claim 8 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:1.

38. The method according to claim 8 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:2.

39. The composition according to claim 19 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:1.

40. The composition according to Claim 19 wherein X₂ consists of an amino acid sequence comprising SEQ ID NO:2.